



## Use of fly screens to reduce *Campylobacter* spp. introduction in broiler houses

Hald, Birthe; Sommer, Helle Mølgaard; Skovgaard, Henrik

*Published in:*  
Emerging Infectious Diseases

*Publication date:*  
2007

*Document Version*  
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

*Citation (APA):*  
Hald, B., Sommer, H. M., & Skovgaard, H. (2007). Use of fly screens to reduce *Campylobacter* spp. introduction in broiler houses. *Emerging Infectious Diseases*, 13(12), 1951-1953.

---

### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

# Use of Fly Screens to Reduce *Campylobacter* spp. Introduction in Broiler Houses

Birthe Hald,\* Helle M. Sommer,†  
and Henrik Skovgård‡

Fly screens that prevented influx of flies in 20 broiler houses during the summer of 2006 in Denmark caused a decrease in *Campylobacter* spp.–positive flocks from 51.4% in control houses to 15.4% in case houses. A proportional reduction in the incidence of chicken-borne campylobacteriosis can be expected by comprehensive intervention against flies in broiler production houses.

*Campylobacteriosis* is a severe gastroenteric human disease of global significance. The incidence correlates with the prevalence of thermophilic *Campylobacter* spp., predominantly *C. jejuni* and *C. coli* (1), in chickens and follows a seasonal cycle in temperate climates for reasons not fully elucidated. The number of cases is lowest in winter and highest in summer (2). In Denmark, the prevalence of *Campylobacter* spp.–infected chicken flocks peaked at 60%–80% in recent summers (3). The population size of flies displays a similar cycle (4). Flies, in particular the house fly, *Musca domestica*, are well-known vectors of several enteric bacterial diseases (5) and are known to carry *Campylobacter* spp. (6–10). Vector flies can transmit *Campylobacter* spp. from outside farm livestock to broiler flocks because large numbers of flies may enter broiler houses by ventilation air (7,11). Our aim was to evaluate the effect of insect screens in addition to existing biosecurity measures against *Campylobacter* spp. infection of broiler chickens in summer.

## The Study

Potential study sites were identified in the Danish Poultry Council's national surveillance database (3) on the basis of the number of *Campylobacter* spp.–positive flocks produced in broiler houses during 2003–2005. All farms practiced hygiene procedures such as separating clean and dirty zones, changing footwear and clothes, and washing hands with disinfecting soap before entering the broiler room. Furthermore, a 3-m zone with short-cut grass

or gravel surrounded the houses. Houses were emptied, cleaned, and dried before each new flock of chickens was brought in. All farmers were instructed to maintain biosecurity and management routines as before the study. Case and control groups were assigned to match each other in *Campylobacter* spp. prevalence and were composed so that the distribution of previous *Campylobacter* spp. prevalence of flocks for each group (June to November during 2003–2005) were equal (Figure 1) and with similar distribution in the presence of other livestock in a periphery of 1.5 km around the farms. Farmers consented to participate before study groups were composed.

According to data from the national Danish *Campylobacter* surveillance program (3), the historical *Campylobacter* spp. prevalence at slaughter during 2003–2005 from June to November had been 51.6% (95/184) (95% confidence interval [CI] 44.3%–59.0%) in case houses and 51.7% (123/238) (95% CI 45.2%–58.2%) in control houses. Thus, before the study, the baseline prevalence for houses in the case and control groups were not significantly different from each other ( $p = 0.99$  by  $\chi^2$  test).

Twenty houses on 11 farms in Jutland, Denmark, were equipped with fly screens by June 1, 2006 (photographs available from [www.vet.dtu.dk/default.aspx?id=20832](http://www.vet.dtu.dk/default.aspx?id=20832)). Fifty-two broiler flocks stocked in the houses after June 1 constituted the cases; the last flock was slaughtered on November 6, 2006. Controls were 70 broiler flocks reared in 25 matched broiler houses on 13 other farms without fly screens; the last flock was slaughtered on November 13, 2006. All houses were ventilated through wall inlets in the long sides of the houses, air outlets through chimneys in the roofs, and gable fans. The study design was based on experience gained in a pilot study in 2004 (11) of 5 farms with parallel case and control houses on each farm. The pilot study showed a significant delay of *Campylobacter* spp. introduction in case houses. However, only a 37% reduction in positive broiler flocks was obtained at slaughter due

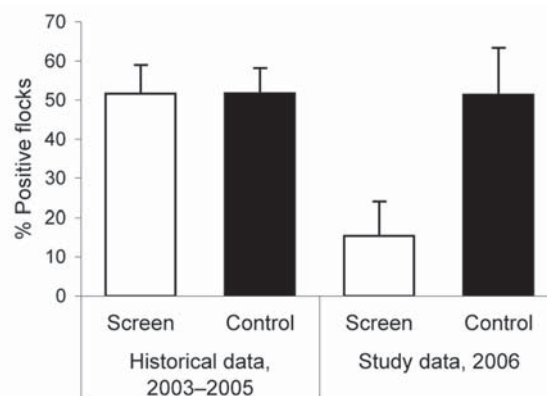


Figure 1. Percentage of *Campylobacter* spp.–positive broiler flocks produced in fly screen houses and control houses June 1 to November 13 during 2003–2005 (historical data) and in 2006 (during intervention). Error bars indicate standard deviation.

\*Technical University of Denmark, Aarhus, Denmark; †Technical University of Denmark, Søborg, Denmark; and ‡University of Aarhus, Lyngby, Denmark

to transmission of *Campylobacter* spp. from control houses to the corresponding case houses.

Broiler flocks were sampled at days 21, 28, and 35. Boots with over-shoe covers were used to walk through the broiler rooms. The over-shoe covers (photographs available from [www.vet.dtu.dk/default.aspx?id=21756](http://www.vet.dtu.dk/default.aspx?id=21756)) were analyzed for *Campylobacter* spp. Results are shown in Table 1. Flocks were slaughtered between days 35 and 42 and sampled by collection of 10 cloacal swabs per flock at the abattoir. Results of the current national surveillance program of *Campylobacter* spp. in broiler production were included in the study as reference to ordinary Danish broiler production. All samples were analyzed by PCR (DANAK [The Danish Accreditation and Metrology Fund] accredited method) detecting thermophilic *Campylobacter* spp. (12).

In fly screen houses (case houses), 15.4% (95% CI 7.7%–27.8%) of the flocks reared during the study period were *Campylobacter* spp. positive at slaughter, whereas the prevalence in *Campylobacter* spp.–positive flocks reared in the control houses was 51.4% (95% CI 40.0%–62.7%). The prevalence in the control houses remained unchanged ( $p = 0.68$  by  $\chi^2$  test) compared with the historical prevalence during June–November, 2003–2005. Figure 1 shows the *Campylobacter* spp. prevalence of flocks from the study with the historical data. The average flock *Campylobacter* spp. prevalence per month in 2006 in fly screen houses and in control houses is shown in Figure 2 with the results of the national Danish *Campylobacter* surveillance program of 1,504 broiler flocks slaughtered in Denmark in specific months.

Data were analyzed with SAS software (SAS Institute, Cary, NC, USA) in the SAS procedure proc genmod with a logit link function and a repeated statement where subject = flock. The repeated statement accounts for the intra-class correlation. In the model, the effects of the fly screen “Screen” of the time between 21 and 35 days “Time”, the interaction “Screen Time” and the effect of the average monthly prevalence level at slaughter “Month” (analyzed as regressor) were analyzed. The status at day 35 was chosen in the analysis instead of the results at slaughter to avoid biases in data due to the increased risk of introducing *Campylobacter* spp. in those flocks slaughtered later and during depopulation and transportation to slaughter. Only 4 flocks were slaughtered during November and were merged with the October flocks in the analysis.

The analysis shows a clear effect of fly screens ( $p = 0.0002$ ) by either complete prevention of infection or by

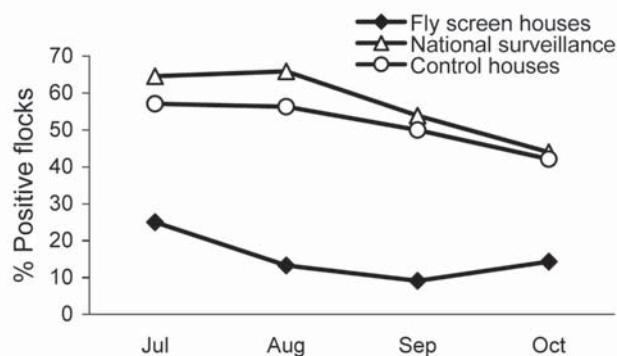


Figure 2. Prevalence per month of *Campylobacter* spp.–positive broiler flocks during the study period (June 1–November 13, 2006) in fly screen houses (52 flocks) and control houses (70 flocks), and the national flock *Campylobacter* spp. prevalence at slaughter of 1,504 flocks according to surveillance data for the same period.

a significant ( $p < 0.0001$ ) delay in onset of infection of the broiler flocks. Results of analyzed sources and estimates of flock *Campylobacter* spp. status in fly screened and in unprotected houses at days 21 and 35 predicted by the applied statistical model are shown in Table 2.

## Conclusions

We showed that preventing flies from entering broiler houses in the summer of 2006 caused a drop in prevalence of *Campylobacter* spp.–positive flocks at slaughter from 51.4% in control houses to 15.4% in case houses. It seems reasonable that the main results found in this study can be extrapolated to the national situation because the selected control houses had a prevalence similar to the national prevalence level for the same period (Figure 2). Installation of effective fly screens in broiler houses in Denmark would most likely decrease the average yearly *Campylobacter* spp. prevalence, and show a major decrease in the summer peak. Presumably, the risk for infection from eating chicken, the main cause of campylobacteriosis in Denmark (13), would be reduced. The expected effect on the incidence of chicken-borne campylobacteriosis has been calculated by Rosenquist et al. (14) to be proportional to the decline in flock *Campylobacter* spp. prevalence.

Our study provides evidence that flies are vectors for *Campylobacter* spp. in broilers and furthermore, probably explains the seasonal variation of *Campylobacter* spp. in

Table 1. *Campylobacter* spp. positive and negative flocks by type of house

Type of house	Day 21		Day 28		Day 35	
	No. positive (%)	No. negative	No. positive (%)	No. negative	No. positive (%)	No. negative
Fly screened (n = 52)	3 (5.8)	49	3 (5.8)	49	4 (7.7)	48
Control (n = 70)	8 (11.4)	62	20 (28.6)	50	30 (45.5)	36

Table 2. Results of analyzed sources and estimates of flock *Campylobacter* spp. status from the applied statistical model

Type of result	p value
Source of variation*	
Screen	0.0002
Time (day of rotation)	<0.0001
Screen time	0.07
Month	0.80
Predicted prevalence of <i>Campylobacter</i> spp.—positive flocks (day 21/35), %	
Fly screen houses	3/ 11
Houses without fly screens	14/ 42

\*Analysis of variance, type 3 test. Significant effects if  $p < 0.05$ .

chicken products. Flies may also play a role in direct transmission of *Campylobacter* spp. to humans (14,15). Certainly, the issue deserves further scientific investigation.

### Acknowledgments

We thank Henrik Bunkenborg, Niels Balle, Allan Balle, Tommy H. Krogh, Niels Borre, Palle Vinstrup, Jacob R. Pedersen, Jan Hedemand; the broiler farmers (Sejer Grimstrup, Jens and Jørgen Black, Per Villumsen, Ole Larsen, Erik and Flemming Hjorth, Karin Lieder, Søren Arne Bruun, Henrik Gammelgård, Jens Theilm, and Jørgen Nørgård) for their participation in the study; Anita Fogh Hansen and Lotte Christensen for laboratory work; and Flemming Bager for his dedicated and constructive criticism of the manuscript.

This project was supported by grant 66032–0128 from The Directorate for Food, Fisheries and Agri Business.

Dr Hald is a veterinarian and member of the *Campylobacter* research group at the National Veterinary Institute in Aarhus. Her main research interest is the epidemiology of *Campylobacter* in poultry, pets, and wildlife.

### References

- Newell DG, Fearnley C. Sources of *Campylobacter* colonization in broiler chickens. *Appl Environ Microbiol*. 2003;69:4343–51.
- Nylen G, Dunstan F, Palmer SR, Andersson Y, Bager F, Cowden J, et al. The seasonal distribution of *Campylobacter* infection in nine European countries and New Zealand. *Epidemiol Infect*. 2002;128:383–90.
- Danish Poultry Council. Database. Danish Meat Association, Copenhagen, Denmark. 2006.
- Nichols GL. Fly transmission of *Campylobacter*. *Emerg Infect Dis*. 2005;11:361–4.
- Busvine JR. Disease transmission by insects: its discovery and 90 years of effort to prevent it. Berlin/Heidelberg: Springer-Verlag; 1993.
- Gregory E, Barnhart H, Dreesen DW, Stern NJ, Corn JL. Epidemiological study of *Campylobacter* spp. in broilers: source, time of colonization, and prevalence. *Avian Dis*. 1997;41:890–8.
- Hald B, Skovgård H, Bang DD, Pedersen K, Dybdahl J, Jespersen JB, et al. Flies and *Campylobacter* infection of broiler flocks. *Emerg Infect Dis*. 2004;10:1490–2.
- Rosel O, Kapperud G. House flies (*Musca domestica*) as possible vectors of *Campylobacter fetus* subsp. *jejuni*. *Appl Environ Microbiol*. 1983;45:381–3.
- Shane SM, Montrose MS, Harrington KS. Transmission of *Campylobacter jejuni* by the housefly (*Musca domestica*). *Avian Dis*. 1985;29:384–91.
- Szalanski AL, Owens CB, McKay T, Steelman CD. Detection of *Campylobacter* and *Escherichia coli* O157:H7 from filth flies by polymerase chain reaction. *Med Vet Entomol*. 2004;18:241–6.
- Hald B, Skovgård H, Pedersen K, Bunkenborg H, Madsen M. Insect screen against *Campylobacter*, an intervention study in broiler houses. In: Abstracts of Scientific Presentation, 13th International Workshop on *Campylobacter*, *Helicobacter* and Related Organisms; Gold Coast, Queensland, Australia; September 4–8, 2005.
- Lund M, Wedderkopp A, Waino M, Nordentoft S, Bang DD, Pedersen K, et al. Evaluation of PCR for detection of *Campylobacter* in a national broiler surveillance programme in Denmark. *J Appl Microbiol*. 2003;94:929–35.
- Wingstrand A, Neimann J, Engberg J, Nielsen EM, Gerner-Smidt P, Wegener HC, et al. Fresh chicken as main risk factor for campylobacteriosis, Denmark. *Emerg Infect Dis*. 2006;12:280–5.
- Rosenquist H, Nielsen NL, Sommer HM, Nørrung B, Christensen BB. Quantitative risk assessment of human campylobacteriosis associated with thermophilic *Campylobacter* species in chickens. *Int J Food Microbiol*. 2003;83:87–103.
- Nelson W, Harris B. Flies, fingers, fomites, and food. *Campylobacteriosis in New Zealand, food-associated rather than food-borne*. *N Z Med J*. 2006;119:U2128.

Address for correspondence: Birthe Hald, National Veterinary Institute, Technical University of Denmark, Høngvej 2, DK-8200, Aarhus N, Denmark; email: bha@vet.dtu.dk

All material published in Emerging Infectious Diseases is in the public domain and may be used and reprinted without special permission; proper citation, however, is required.

EMERGING INFECTIOUS DISEASES *online*

[www.cdc.gov/eid](http://www.cdc.gov/eid)

To receive tables of contents of new issues send an email to [listserv@cdc.gov](mailto:listserv@cdc.gov) with subscribe eid-toc in the body of your message.